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Journal of Applied Sciences and Environmental Management

WORLD BANK ASSISTED NATIONAL AGRICULTURAL RESEARCH PROJECT (NARP) - UNIVERSITY OF PORT HARCOURT ISSN: 1119-8362

Vol. 13, Num. 2, 2009, pp. 69-74

Journal of Applied Sciences & Environmental Management, Vol. 13, No. 2, June, 2009, pp. 69-74

Camphor: risks and benefits of a widely used natural product

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Code Number: ja09027

ABSTRACT

This study analyzes the main aspects of the non-clinical profile of D-Camphor, a natural product widely used as a common remedy for several symptoms. The pharmacology, pharmacokinetics and toxicity of this substance are analyzed, with regard to all the literature available, in order to assess a risk profile and better understand the positive and negative aspects connected with its use. The general conclusion is that the main risks of camphor as a medicinal product are mainly due to a somehow diffused attitude of considering it as "not a real medicine", and to its consequent sometimes not sufficiently careful administration. @ JASEM

Camphor is a natural product derived from the wood of the camphor laurel (Cinnamonum camphora L.) trees through steam distillation and purification by sublimination; the trees used should be at least 50 years old. It also occurs in some other related trees in the laurel family, notably Ocotea usambarensis Eng., and can also be obtained from the plant Lippia dulcis Trev., but this is not a major industrial source (Compadre et al., 1986). A major source of camphor in Asia is Ocimum kilimandscharicum Baker ex Gurke. Camphor can also be produced synthetically from vinyl chloride and cyclopentadiene, passing through the intermediate dehydronorbornyl chloride. The naturally occurring form is dextroortatory and the synthetic form optically inactive (Budavari, 1989; Reynolds, 1989). Camphor has a counter-irritant, rubefacient and mild analgesic action, and is a major component of liniments for relief of fibrositis, neuralgia and similar conditions. It can be used as a mild expectorant; when ingested camphor has irritant and carminative properties. Camphorated-oil, a solution in oil given through intramuscular or subcutaneous way, can be used as a circulatory and respiratory stimulant, but this use is considered hazardous. When in combination with menthol and chenodeoxycholic acid it has been used to aid dispersal of bile duct stones, although this is no longer recommended (Reynolds, 1989). Aim of the present work is to provide an overview over pharmacological, pharmacokinetical and toxicological aspects of camphor, in order to assess its safety profile and evaluate the eventual level of risk connected with its use.

PHARMACOLOGY

Camphor, a natural product derived from the wood of the tree Cinnamomum camphora, has a long history of use as antiseptic, analgesic, antipruritic, counterirritant and rubefacient (Ellenhorn and barceloux, 1998; Liebelt and Shannon, 1993). Its success and wide medical use, especially in topical preparations, is connected to its mild local anesthesizing effect and to the production of a circumscribed sensation of heat, together with its characteristic and penetrating odour that is by most of people associated to the idea of a strong and effective medicine (Gibson et al., 1989). Camphor is today mostly used in the form of inhalants and of camphorated oil, a preparation of 19% or 20% camphor in a carrier oil, for the home treatment of colds (Jochen and Theis, 1995) and as a major active ingredient of liniments and balms used as topical analgesics (Xu et al., 2005). The antitussive, nasal decongestant and expectorant action of camphor and of its derivatives was one of the first ones to be systematically investigated (Inoue and Takeuchi, 1969). Its nasal decongesting activity seems to be not purely mechanic, but connected with the stimulation of cold receptors in the nose. The inhalation of camphor vapours (so as the one of eucalyptus and menthol vapours) on a sample of volunteers increased the nasal sensation of airflow through the induction of cold sensation in the nose, despite of actually not affecting nasal resistance to airflow (Buroow et al., 1993). Camphor was administered in the form of aromatic vapor, at the concentrations of 50, 133 and 500 µg Γ^1 , to guinea pigs subject to chemically induced cough. No effect were registered at the lowest concentrations, but 500 µg I⁻¹ camphor gave a 33% reduction of cough frequency, to which an increase in cough latency coincided (Laude et al., 1994). The analgesic proprieties of camphor are largely known and applied, but little is known about the molecular mechanisms that are at their basis. (Xu et al., 2005). Mogrich et al. (2005) demonstrated that camphor activates TRPV3, a member of transient receptor channel superfamily, leading to excitation and desensitization of sensory nerves. The notorious effect of generation of a sensation of heath associated with topic application of camphor (Green, 1990) is a consequence of this activation. Anyway excessive and repeated application of camphor can lead to sensibilization of TRPV3, in apparent contrast with its analgesic role (Moqrich et al., 2005; Peier et al., 2002). The antipruritic, analgesic and counterirritant activity of camphor is instead associated with its capacity of activating TRPV1- another member of TRP channel superfamily -at the level of dorsal root gangliar [DRG] neurons and inhibiting TRPA1 channels (Moqrich et al., 2005; Nagata et al., 2005), action that is in common with other TRPV1 agonists (Koplas et al., 1997; Bhave et al., 2002). Capsaicin shares the same action with camphor, but performs it more slowly and less completely; on the other side camphor efficacy is lower, since higher concentrations are required (Xu et al., 2005). Studies on rats demonstrated that the actions of capsaicin and camphor are segregated [8], i.e. they are mediated by distinct channel regions, and camphor did not activate TRPV1 in capsaicininsensitive chickens (Xu et al., 2005, Jordt and Julius, 2002). Camphor also inhibits other related TRP channels such as ankyrin-repeat TRP1 (TRPA1), which is a further evidence underlying its analgesic effects (Xu et al., 2005). Camphor can modulate the activities of hepatic enzymes involved in phase I and phase II drug metabolism. 50, 150 and 300 mg Kg⁻¹ of camphor dissolved in 0.1 ml of olive oil was administered daily to female Swiss Albino mice during 20 days. At its highest concentration it caused a significant increase in the activities of cytochrome P450, cytochrome b5, aryl-hydrocarbon hydroxylase and glutathione S-transferase, significantly elevating the level of reduced glutathione in the liver (Banerjee et al., 1995). Camphor was shown to inhibit mitochondrial respiration. Administration of up to 8 μ M of camphor inhibited respiration rate in rat-liver mitochondria, nearly halving the oxygen consumption; this suggests that camphor may be used in oxygenating tumors prior to radiotherapy (Guilland-Cumming and Smith, 1979; 1982). Camphor can also be a potential radiosensitizing agent in radiotherapy. Treatment with camphor (0.5 µmol · body wt⁻¹) 45 minutes before local xirradiation at the dose levels of 30, 80, 100 or 120 Gy was performed on male C3H/Jax mice bearing transplanted mammary tumours. Sequential measurement of the tumour volumes during 45 days after the irradiation revealed a 4.8 delay of the maximum enhancement ratios in tumour growth (Goel and Roa, 1988). D-camphor (1100 μ g ml⁻¹) inhibited oxidative metabolism in E.coli (Cardullo and Gilroy, 1975). Succinic, lactic and NADH-oxidase activities were inhibited, while NADH and succinic DCPIP oxidoreductase enzymes were unaffected. The restoration of succinic oxidase activity by ubiquinone (Q6) but not by vitamin K1 indicates that D-camphor may operate this inhibition by affecting quinone functions.

Interactions

Very few studies of pharmacological interactions between camphor and other compounds are present in literature. In a study combining the administration of D-camphor and an extract from fresh crataegus berries, a synergic action of the two preparations emerged in ameliorating cardiac performances. Both D-camphor and the extract contributed in an increase in total peripheral resistance induced by an increase tone of the arterioles, and while the former appeared to be the main factor in inducing the rapid initial effect, the former added a long-lasting effect (Belz and Loew, 2003).

PHARMACOKINETICS

Camphor is readily absorbed from all the sites of administration, after inhalation, ingestion or dermal exposure (Baselt and Cravey, 1990). Peak plasma levels were reached by 3 hours post-ingestion when 200 mg camphor was taken alone, and 1 hour postingestion when it was ingested with a solvent (Tween 80) (Koppel et al., 1988). In case of dermal application, the volume of the absorption is relatively low in comparison with the speed of the process. After application of different numbers of commercial patches (2, 4 or 8) to the skin of human subjects during 8 hours, the levels of camphor in the plasma were assayed with selective gascromatography (Maetin et al., 2004; Valdez et al., 1999). Maximum camphor plasma concentration resulted in a range between 35.2 and 46.8 ng ml-1 in the case of 8 patches, between 19.6 and 34 ng ml-1 for the 4 patches while almost undetectable concentrations were observed when only 2 patches had been applied, showing that dermal absorption is prompt but not massive. Camphor is distributed throughout the whole body, and can permeate the placenta; for this reason it must be recommended that the use of this product is avoided during pregnancy and lactation (Sweetman, 2005). Its volume of distribution is 2- 4 L/kg (Koppel et al., 1988); plasma protein binding has been estimated as 61% (Koppel et al., 1982). After its absorption and distribution, camphor undergoes hepatic metabolises: it is hydroxylated in the liver into hydrocycamphor metabolites (Sweetman, 2005). Asahina and Ishidate (1933; 1934; 1935) isolated cis-and trans-rthydroxycamphor (20%) from the urine of dogs, and 5-hydroxycamphor [as major metabolite] and 3hydroxycamphor from the urine of rabbits (Shimamoto, 1934). Robertson and Hussain (1969) observed that (+)-camphor and (-)-camphor increase the content of glucuronide in the urine of rabbits (+)-camphor was moreover reduced to (+)-borneol as well as being hydroxylated to (+)-5-endo-hydroxycamphor. Hydroxylamphor, as well as norcamphor, pericyclocamphanone and 5,5-difluoroca

performed by Cytochrome P-450 (Collins and Loew, 1988), a class of heme-containing monooxygenases that are distributed in the whole body (Boxenbaum, 1984), by hydrogen abstraction (Wand and Thompson, 1986). Cytochrome P-450 is responsible for camphor conversion into 5hydroxycamphor (Gelb et al., 1982), while 3hydroxycamphor is the primary product of nonenzymatic hydroxylation of camphor (Land and Swallow, 1979). Camphor hydroxylation by cytochrome P-450 occurs with a different regionspecificity for camphor and its related compounds (Collins and Loew, 1986). Hydroxylated metabolites are then conjugated with glucuronic acid and excreted in the urine (Sweetman, 2005). The half-life of 200 mg of camphor was 167 minutes when ingested alone, and 93 minutes when ingested with a solvent (Tween 80) (Koppel et al., 1988).

TOXICITY

Camphor occurs in nature in its dextrorotatory form (D-camphor), while the laevorotatory form (Lcamphor) exists only as a synthetic form. The two enantiomers present different profiles of toxicity. D-camphor, L-camphor and their racemic mixture were tested for toxicity in mice. At 100 mg · Kg b.w.⁻¹ the natural form was non toxic, while the synthetic form induced different kinds of toxic and behavioural effects such as body jerks and hunched posture; the racemic mixture showed similar effects to the L-form (Chatterjie and Alexander, 1986). The oral administration of acute doses of Dcamphor to rats and rabbits caused pronounced signs of toxicity. In rats, the consume of food was reduced proportionally to the administered dose, starting from 464 mg · Kg b.w.⁻¹ · day⁻¹, and at 1000 mg · Kg b.w.⁻¹ · day⁻¹ convulsions and piloerection were observed, connected with a reduction of motility and weight gain. Reduced body weight gain and food consumption were observed in rabbits treated with 681 mg · Kg b.w.¹ · day⁻¹ (Leuschner, 1997). Camphor showed porphyrogenic activity in primary cultures of chick embryo - liver cells, with enhanced porphyrin accumulation ranging from 5- to 20-fold (Bonkovsky et al., 1992). The main problems about camphor toxicity in humans are connected more to the large availability of camphor-containing products and their diffused perception as unhazardous medicines rather than in the intrinsic toxicity of camphor. The daily maximum human therapeutic dose is in fact approximately 1.43 mg · Kg⁻¹, which corresponds to a therapeutic ratio of more than 450 for the endpoint toxicity, reflecting a wide margin of safety (Leuschner, 1997). On the other side, as mentioned above, camphor is present in several over-the-counter products, its use as a familiar remedy is commonly accepted, but still some lack of information persists among the consumers. Cases of camphor intoxication in humans, especially children, are relatively frequent, mostly because of accidental ingestion (Siegel and Wason, 1986). More than 100000 cases of ingestion exposures to camphor-containing products were registered between 1990 and 2003 (Manoguerra et al., 2006), causing a range of symptoms that comprises convulsion, lethargy, ataxia, severe nausea, vomiting and coma (Koppel et al., 1988; Manoguerra et al., 2006).

Reproduction toxicity

D-camphor was orally administered to pregnant rats and rabbits during the period of organogenesis to test its embryotoxicity. Doses up to 1000 mg · Kg b.w.⁻¹ · day⁻¹ to rats and up to 681 mg · Kg b.w.¹ · day⁻¹ to rabbits showed no teratogenic effects, and in none of the animals were observed higher rates of mutations or malformations (Leuschner, 1997).

Mutagenicity and cancerogenicity

In a Salmonella/microsome assay, the upper limit of the dose interval tested for (+/-) camphor resulted to be the highest non-toxic dose, suggesting that the compound is not mutagenic in the Ames test (Gomes-Carneiro et al., 1998). A single dose of camphor (0.5μ M · g⁻¹) administered 30, 45 or 60 minutes before gamma irradiation significantly reduced the frequency of sisterchromatid exchanges in mouse bone marrow, showing therefore a radiomodifying influence (Goel et al., 1989).

CONCLUSIONS

Camphor is familiar to many people as a principal ingredient in topical home remedies for a wide range of symptoms, and its use is well consolidated among the population of the whole world, having a long tradition of use as antiseptic, antipruritic, rubefacient, abortifacient, aphrodisiac, contraceptive and lactation suppressant. This compound has also a long history of scientific studies on its action and on the way through which it is metabolized in the organisms of both humans and animals, due to the general interest that it has always arisen among common people and scientists. Already in 1879, Schmiedeberg and Meyer were analyzing the metabolites isolated from the urine of dogs that had been fed with (+/-) camphor (Schmiedeberg and Meyer, 1879), and during the first half of the twentieth century the number of studies focused on its pharmacology and pharmacokinetics has been remarkable. The bibliographic search that was performed for the compilation of this toxico-pharmacological overview revealed a rich literature existing on camphor, and put in evidence the large amount of works focused on toxic aspects of camphor that were published during the last 30 years; a great number of reports concerning cases of camphor intoxication were also collected. In most cases camptor intoxication occurred following accidental ingestion of camphor-containing product, and sometimes lethal episodes of intoxication of infants due to application of camphor to their nostrils were collected. As it emerges from all the observed data the toxic risks of camphor-containing products in general, and of camphorated oil in particular, are connected essentially with its improper uses, e.g. accidental ingestion, but camphor does not represent a threaten for safety when used on the target patients, following the indicated dosages and the contraindications. Special care must be taken during pregnancy, due to the fact that camphor crosses the placental barrier, and camphor and camphor containing products should be avoided in children who have a history of febrile convulsions or other predisposing factors for convulsions (Galland et al., 1992).

In the past, when camphor was used medicinally, the oral doses ranged from 120-300 mg (Wade, 1977), and the parenteral dose range was from 60200 mg (not recommended any more). Camphorated oil can be used with no risks for safety when following the prescriptions. The relatively diffused tendency to the improper use of camphor [high dosages, accidental ingestion, use on infants] is connected with the perception of the product, by many consumers, as a sort of "panacea" with no contraindication. More and more accessible information is therefore necessary to bring to a "responsabilization" of the consume of this product, in order to avoid hazardous situations. All the above considerations allow the conclusion that camphor in its form of camphorated oil can be safely used at the proposed dosages, on the indicated patients target, for topic application.

REFERENCES

- Asahina, Y Ishidate, M (1933). Ber. dtsch. chem. Ges., 67: 71.
- Asahina, Y Ishidate, M (1934). Ber. dtsch. chem. Ges., 68B: 967. Asahina, Y Ishidate, M (1935). Ber. dtsch. chem. Ges., 68B: 967. Banerjee, S, Welsch CW Rao, AR (1995). Modulatory influence of camphor on the activities of hepatic carcinogen metabolizing enzymes and the levels of hepatic and
- extrahepatic reduced glutathione in mice. Cancer Letters 88(2): 163-169.
- Baselt, RC; Cravey, RH (1990). Disposition of toxic drugs and chemicals and drugs 3rd ed. Year Book Medical Publishers Inc.
- Belz, GG; Loew, D (2003). Dose-response related efficacy in orthostatic hypotension of a fixed combination of D-camphor and an extract from fresh crataegus berries and the
- back, G, Zou, W; Wang, H; Brasier, D; Oxford, GS; Gereau, RW (2002). cAMP-dependent protein kinase regulates desensitization of the capsaicin receptor (VR1) by direct phosphorylation. Neuron 35: 721–731.
- Bonkovsky, HL; Cable, EE; Cable, JW; Donohue, SE; White, EC; Greene, YJ; Lambrecht, RW; Srivastava, KK; Arnold, WN (1992). Porphyrogenic properties of the terpenes camphor, pinene, and thujone (with a note on historic implications for absinthe and the illness of Vincent van Gogh). Biochemical Pharmacol 43: 2359-2368. Boxenbaum, H, (1984). Interspecies Pharmacokinetic Scaling and the Evolutionary-Comparative Paradigm. Drug Metab Rev 15 1071-1121.
- Budavari, S (Ed.), (1989). The Merck Index. 11th Edition. Merck and Co Inc, Rahway, USA.
- Burrow, A; Eccles, R; Jones, AS (1983). The effects of camphor, eucalyptus and menthol vapour on nasal resistance to airflow and nasal sensation. Acta Otolaryngol 96: 157-161
- Cardullo, MA; Gilroy, JJ (1975). Inhibition of oxidative metabolism in Escherichia coli by dcamphor and restoration of oxidase activity by quinones. Can J Microbiol 21: 1357-• 1361

- Chatterjie, N; Alexander, GJ (1986). Anticonvulsant properties of spirohydantoins derived from optical isomers of camphor. Neurochem Res 11: 1669-1676. Collins, JR; Loew, GH (1988). Theoretical study of the product specificity in the hydroxylation of camphor, norcamphor, 5,5-difluorocamphor, and pericyclocamphanone by cytochrome P450cam. J Biol Chem 263: 3164-3170. Compadre, CM; Robbins, E; Kinghorn, AD (1986). The intensely sweet herb, Lippia dulcis Trev.: historical uses, field enquiries, and constituents. J Ethnopharmacol 15: 89-106. Ellenhorn, M.J.; Barceloux, D.G. (1998). Camphor. In Medical Technology: Diagnosis and treatment of human poisoning. New York: Elsevier, pp.: 505-507.
- Galland, MC; Griguer, Y; Morange-Sala, S; Jean-Pastor, MJ; Rodor, F; Jouglard, J (1992). Convulsions febriles: faut-il contre-indiquer certains medicaments? Therapie 47: 409-• 414
- Gelb, MH; Heimbrook, DC; Miilkonen, P; Sligar, SG (1982). Stereochemistry and deuterium isotope effects in camphor hydroxylation by the cytochrome P450cam Gool, HC; Roa, AR (1988). Radiosensitizing effect of camphor on transplantable mammary adenocarcinoma in mice. Cancer Letters 43: 21-27.

- Goel, HC; Singh, S; Singh, SP (1989). Radiomodifying influence of camphor on sister-chromatid exchange induction in mouse bone marrow. Mutation Res, 224: 157-160. Gomes-Carneiro, MR; Felzenszwalb, I; Paumgartten, FJ (1998). Mutagenicity testing (+/-)-camphor, 1,8-cineole, citral, citronellal, ()-menthol and terpineol with the Salmonella/microsome assay. Mutation Res 416: 129-136.
- Green, BG (1990). Sensory characteristics of camphor. J Invest Dermatol 94: 662-666. Guilland-Cumming, D; Smith, GJ (1979). Mitochondrial respiration depressed by camphor: a possible aid in radiotherapy. Experientia, 35: 659. Guilland-Cumming, DF; Smith, GJ (1982). The effect of camphor on mitochondrial respiration. Experientia, 38: 236-237.

- Guilland-Cumming, DF; Smith, GJ (1982). The effect of camphor on mitoconordinal respiration. Experientia, 38: 230-237. Inoue, Y; Takeuchi, S (1969). Expectorant-like action of camphor derivatives. Nippon Ika Daigaku Zasshi, 36: 351-354. Jochen, GW; Theis, MD (1995). Camphorated oil: still endangering the lives of Canadian children. Can Med Ass J 152: 1821-1824. Jordt, SE; Julius, D (2002). Molecular basis for species-specific sensitivity to "hot" chili peppers. Cell 108: 421– 430. Koplas, PA, Rosenberg; RL; Oxford, GS (1997). The role of calcium in the desensitization of capsaicin responses in rat dorsal root ganglion neurons. J Neurosci 17: 3525–3537. Koppel, C; Martens, F; Schirop; Th; Ibe, K (1988). Hemoperfusion in acute camphor poisoning. Int Care Med 14: 431-433. Koppel, C; Tenczer, J; Schirop, T; Ibe, K (1982). Camphor poisoning -babuse of camphor as a stimulant. Archiv Toxicol 51: 101-106.
- Land, EJ; Swallow, AJ (1979). Some free radical reactions of camphor in relation to the action of cytochrome P450. J Chem Soc, Faraday Trans I 75: 1849-1856.

- Laude, EA, Morice, AH; Grattan, TJ (1994). The antitussive effects of menthol, camphor and cineole in conscious guinea-pigs. Pulm Pharmacol 7: 179-184.
 Leuschner, J (1997). Reproductive toxicity studies of D-camphor in rats and rabbits. Arzneimittelforschung, 47: 124-128.
- •

- Leuschner, J (1997). Heproductive toxicity studies of D-camphor in rats and rabbits. Arzneimittelforschung, 47: 124-128. Liebelt, EL; Shannon, MW (1993). Small doses, big problems: a selected review of highly toxic common medications. Ped Emerg Care 9: 292-297. Manoguerra, AS; Erdman, AR; Wax, PM; Nelson, LS; Caravati, EM; Cobaugh, DJ; Chyka, PA; Olson, KR; Booze, LL; Woolf, AD; Keyes, DC; Christianson, G; Scharman, EJ; Troutman, WG (2006). Camphor Poisoning: an evidencebased practice guideline for out-of-hospital management. Clin Toxicol (Phila) 44: 357-370 Martin, D; Valdez, JS; Boren, J; Mayersohn, M (2004). Dermal absorption of camphor, menthol, and methyl salicylate in humans. J Clin Pharmacol 44: 1151-1157. Moqrich, A; Hwang, SW; Earley, TJ; Petrus, MJ; Murray, AN; Spencer, K; Andahazy, M; Story, GM; Patapoutian, A. (2005). Impaired thermosensation in mice lacking TRPV3, a heat and camphor sensor in the skin. Science 307:1468 –1472. •
- Nagata, K; Duggan, A; Kumar, G; Garcia-Anoveros, J (2005). Nociceptor and hair cell transducer properties of TRPA1, a channel for pain and hearing. J Neurosci 25: 4052-4061.
- Peier, AM; Reeve, AJ; Andersson, DA; Moqrich, A; Earley, TJ; Hergarden, AC; Story, GM; Colley, S; Hogenesch, JB; McIntyre, P; Bevan, S; Patapoutian, A (2002). A heat-sensitive TRP channel expressed in keratinocytes. Science 296: 2046 –2049. Reynolds, JEF (Ed.), (1989). Martindale The Extra Pharmacopeia. 29th edition. The Pharmaceutical Press, London. Robertson, JS; Hussain, M. (1969). Metabolism of camphors and related compounds. Biochemi J 113: 57-65.
- •

- Hobertson, JS; Hussain, M. (1969). Metabolism of campnors and related compounds. Biochemi J 113: 57-65.
 Schmiedeberg, O; Meyer, H. (1879). Hoppe-Seyl. Z. 3: 422.
 Shimamoto, T, (1934). Sci. Pap. let. Phys. Chem. Res., Tokyo, no. 529, pp. 52-58, 59-62.
 Siegel, E; Wason, S (1986). Camphor toxicity. Ped Clin North Am 33: 375-379.
 Sweetman, S.C. (ed.), (2005). Martindale: The Complete Drug Reference. 34th edition. The Pharmaceutical Press, London. Valdez, JS; Martin, DK; Mayersohn, M (1999). Sensitive and selective as chromatographic methods for the quantitation of camphor, menthol and methyl salicylate from human plasma. J of Chromatogr B: Biomed Sci Appl 729: 163-171. Wade, A (ed.) (1977). Martindale The Extra Pharmacopeia. 27th edition. The Pharmaceutical Press, London.
- Wand, MC, Tompson, JA (1986). Cytochrome P450-catalyzed rearrangement of a peroxyquinol derived from butylated hydroxytoluene. Involvement of radical and cationic intermediates. J Biol Chem 261 14049-14056.
- Xu, H; Blair, NT; Clapham, DE (2005). Camphor activates and strongly desensitizes the transient receptor potential vanilloid subtype 1 channel in a vanilloid-independent mechanism. J Neurosci 25: 8924-8937.

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